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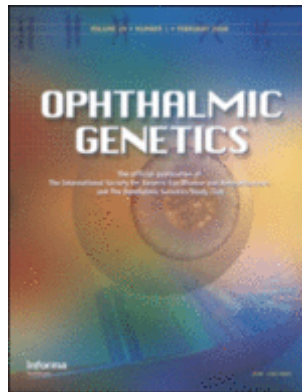
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A genome-wide association study implicates that the *TTC39C* gene is associated with diabetic maculopathy with decreased visual acuity

Ophthalmic Genetics

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ABSTRACT

Background: Diabetic maculopathy is a form of diabetic retinopathy. The visual acuity of one third of patients with diabetic maculopathy will be affected. The purpose of this study was to identify genetic contributors of diabetic maculopathy with decreased visual acuity based on a genome-wide association approach using a well-defined Scottish diabetic cohort.

Methods: We used linked e-health records of diabetic patients to define our cases and controls. The cases in this study were defined as type 2 diabetic patients who had ever been recorded in the linked e-health records as having maculopathy (observable or referable) in at least one eye and whose visual acuity of the eye was recorded to have decreased between the first and the last visual acuity record of that eye in the longitudinal e-health records. The controls were defined as a type 2 diabetic individual who had never been diagnosed with maculopathy or retinopathy in the linked e-health records. Anyone who had laser photocoagulation treatment was also excluded from the controls. A standard genome-wide association approach was applied.

Results: Overall, we identified 469 cases and 1,374 controls within the Genetics of Diabetes Audit and Research in Tayside Scotland (GoDARTS) dataset. We found that the P value of rs9966620 in the *TTC39C* gene was 4.13×10^{-8} , which reached genome-wide significance.

Conclusions: We suggest that the *TTC39C* gene is associated with diabetic maculopathy with decreased visual acuity. This needs to be confirmed by further replication studies and functional studies.

KEYWORDS

Diabetic maculopathy; Visual acuity; Genome-wide association study; *TTC39C*; Genetics;

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44 INTRODUCTION

45 Maculopathy is defined as any pathological condition affecting the macula, a highly sensitive
46 region located centrally on the retina which is responsible for sharp, clear, and accurate colour
47 vision (1). It is commonly associated with old age, eye surgery/trauma and diabetes mellitus (1).
48 Maculopathy commonly causes impaired vision but in severe cases can lead to blindness (2).
49 According to a report by the World Health Organization in 2014, maculopathy was responsible
50 for 3.1% of all visual impairment and 6.6% of all blindness in 2010 (2,3). Between 1990 – 2010,
51 there was an 81% increase in the number of visually impaired people and a 36% increase in
52 numbers of blind people due to maculopathy(2). Maculopathy and its subsequent visual
53 disturbances are linked to many adverse health issues. One such example is falls, which
54 (especially in geriatric patients) can result in fractures, subsequent reduction in quality of life,
55 and an increased number of years spent with disability (4). The economic impact of
56 maculopathy is enormous, with an estimated cost of over £100M in 2010 in England alone,
57 incorporating aspects such as screening, management and home care (5).
58 Diabetic maculopathy (DM) is a type of diabetic retinopathy and it is a major eye complication
59 and visual impairment amongst people with diabetes (2). According to the latest diabetic
60 retinopathy guidelines by The Royal College of Ophthalmologists in the UK, DM can be
61 classified as either focal oedema, diffuse oedema, ischemic, or mixed (6). An epidemiological
62 study of DM in Germany showed a prevalence rate of 15% in type 1 and 23% in type 2 diabetic
63 patients (7). Environmental risk factors of DM include longer duration of diabetes, high glycated
64 haemoglobin (HbA1C) levels, prior high-risk proliferative diabetic retinopathy, presence of

diabetic neuropathy, hypertension, anaemia, elevated serum lipid (triglycerides and cholesterol) levels and raised creatinine levels (7–9).

The genetic mechanism of DM is poorly understood and only limited numbers of genetic studies have been performed, particularly on diabetic macular oedema, a subtype of DM. One of these studies suggested that three polymorphisms in the nitric oxide synthase 3 (*NOS3*) gene were associated with an increased risk of developing diabetic macular oedema (10). The C-634G polymorphism in the vascular endothelial growth factor A (*VEGFA*) gene has also been demonstrated to be associated with development of diabetic macular oedema and diabetic retinopathy, in addition to correlating with macular retina thickness in type 2 diabetics (11). More recently, genetic variations in MicroRNA-146a and the vascular endothelial growth factor C (*VEGFC*) gene were found to be significantly associated with diabetic macular oedema in patients with type 2 diabetes (12,13). Whilst there have been a few genome-wide association studies (GWAS) performed on diabetic retinopathy across various ethnic groups, no GWAS have yet been performed specifically on DM (14–17). Graham PS et al. has performed a GWAS on diabetic macular oedema although no GWAS significance was generated. (18)

Given the fact that people are living longer with diabetes and the increasing prevalence of diabetic eye complications including maculopathy, it is necessary to understand the genetic mechanisms of DM as we seek new ways to relieve the burden of morbidity that DM creates worldwide. Therefore, this study seeks to identify genetic variants for DM using a GWAS approach in a well-defined diabetes cohort within Scotland.

MATERIALS AND METHODS

Participants

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3 88 To identify genetic risk factors for diabetes and its complications, the Genetics of Diabetes Audit
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5 89 and Research in Tayside Scotland (GoDARTS) project was established in 2005. All participants
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8 90 (including diabetic and non-diabetic individuals, mainly Scottish) completed a lifestyle
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10 91 questionnaire, a baseline clinical examination, and provided their biological samples (blood
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12 92 and/or urine). The participants also gave broad consent to allow their health information and
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14
15 93 biological samples to be used for future scientific research purposes. In addition, the
16
17 94 participants gave permission to have their personal information linked to the National Health
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19 95 Service (NHS) medical records anonymously. This information included their personal health
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22 96 status, their general practice clinic visits, outpatient appointments, prescribing history and
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24 97 hospital admissions. Furthermore, their personal information was also anonymously linked with
25
26 98 the Scottish Care Information-Diabetes Collaboration (SCI-DC) database, which is another
27
28 99 electronic health record system designed to track local diabetic patients and help health
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31 100 professionals to provide better health care in Scotland. Further information about the GoDARTS
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33 101 project is available in the public domain at <https://godarts.org>. This study has followed the
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35 102 principles of the Declaration of Helsinki. Ethics approval has been granted by Tayside
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37
38 103 Committee on Medical Research Ethics (REC reference 053/04).

39
40 104 At the time of this study, the GoDARTS project had recruited 9,439 diabetic patients, 6,927 of
41
42 105 which were already genotyped by DNA chips. All GoDARTS participants' health information
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45 106 was anonymously linked with their NHS and SCI-DC medical records from June 1996 until June
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47 107 2011.

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49 108 **Definition of cases and controls**

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51 109 The cases in this study were defined as type 2 diabetic patients who had ever been recorded in
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54 110 the linked e-health records as having maculopathy (observable or referable) in at least one eye
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and whose visual acuity of the eye was recorded to have decreased between the first and the last visual acuity record of that eye in the longitudinal e-health records. The controls were defined as a type 2 diabetic individual who had never been diagnosed with maculopathy or retinopathy in the linked e-health records. Anyone who had laser photocoagulation treatment was also excluded from the controls. A standard genome-wide association approach was applied. The definition of visual acuity is summarized in Supplementary Table S1. The diagnosis of DM and assessment of visual acuity was performed by ophthalmologists within the annual national retinal screening service offered to diabetic patients. The diagnostic criteria for diabetic retinopathy or DM are summarized in the Supplementary Table S2.

Genotyping and quality control

The GoDARTS project used two types of DNA chips to genotype its participants with diabetes. The SNP6.0 (Affymetrix, Santa Clara, CA, USA) chips were used on 3,673 subjects, and the OmniExpress (Illumina, Inc., San Diego, CA, USA) chips were used on 3,254 subjects. The standard genotyping quality-control protocols were established for the above studies (WTCCC2 and SUMMIT) (19,20).

Statistical analysis

The imputation of non-directly genotyped single nucleotide polymorphisms (SNPs) was carried out using SHAPEIT and IMPUTE2 software, with reference files from the 1000 genome phase I datasets (21,22). The cut-off value ($r^2 < 0.3$) recommended by IMPUTE2 was applied to remove poorly imputed SNPs.

Standard quality control steps were applied during data analysis, such as removal of individuals with more than 5% genotype data missing, SNPs with missing genotype of more than 5%, or SNPs with less than 1% minor allele frequency and SNPs that failed Hardy–Weinberg tests ($P <$

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3 134 0.000001). PLINK was used as the primary software for data analysis
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5 135 (<https://www.cog-genomics.org/plink2>) (23). SNPs on sex chromosomes and mitochondria
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7
8 136 were also excluded. Detection of population stratification and removal of population outliers
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10 137 were performed using the multidimensional scaling (MDS) analysis integrated in PLINK. To
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12 138 indicate the level of stratification, a lambda value was generated by MDS. Samples with pi-hat >
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14 139 0.125 were removed due to relatedness. *P* values were calculated by the logistic regression
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16 140 tests integrated in PLINK with covariates of age, sex, body mass index (BMI), cholesterol,
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18 141 triglycerides, high-density lipoprotein (HDL), low-density lipoprotein (LDL) and HbA1c (using the
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22 142 latest available health records). A *P* value of less than 5×10^{-8} was considered to be
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24 143 statistically significant.
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27 144 This study also used many GWAS related software such as FUMA for generating Manhattan
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29 145 plots, LocusZoom for regional visualization, and FUMA for generating a corresponding Q-Q plot
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31 146 to evaluate differences between the cases and controls caused by potential confounders (e.g.
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33 147 different genotyping laboratories, different DNA extraction methods, etc.) (24,25). SPSS 22
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35
36 148 software (IBM Corp., Armonk, NY, USA) was used to compare means of all covariates (except
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38 149 sex) in cases and controls through the independent sample t-test. Sex difference was
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40 150 compared by chi-square test. The whole workflow is shown in Supplementary Figure S1.
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44
45 152 **RESULTS**

46
47 153 After excluding type 1 diabetic samples, population outliers, and related samples from 6,927
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49 154 diabetic patients with genetic information from Affymetrix and Illumina chips, we were left with a
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52 155 study population of 4,852 unrelated type 2 diabetes individuals for further analysis. Among
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54 156 these patients, we identified 1,240 samples with diabetic maculopathy. However, only 469
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participants had a decreased visual acuity in the corresponding eye and were suitable for inclusion as cases. We also identified 1,374 patients as controls, after excluding patients who had diabetic retinopathy without DM, patients who had previous laser treatment, and patients who were already cases. Therefore, there were 469 cases (males=192, females=277) and 1,374 controls (males=630, females=744) for the further association analysis. The prevalence of diabetic maculopathy with decreased visual acuity in our cohort was 25.4% [469/(469+1,374)]. The means of sex, age, BMI, cholesterol, triglycerides, HDL, LDL, HbA1c were compared between cases and controls. There were statistically significant differences in age, triglycerides, HDL and HbA1c between the cases and controls, but no statistically significant difference in sex, BMI, cholesterol and LDL (Table 1).

Affymetrix SNP6.0 chips contained 704,847 directly genotyped and quality-controlled SNPs and Illumina OmniExpress chips contained 601,394 directly genotyped and quality-controlled SNPs. Overall, 6,717,712 genotyped and imputed SNPs were available for association analysis after standard quality control steps of genotyping and imputation. There was no need to further adjust for population stratification since the lambda value was 1.00, indicating a homogeneous population. We then performed logistic regression tests integrated in PLINK adjusting for sex, age, BMI, cholesterol, triglycerides, HDL, LDL, HbA1c. We found that the SNP rs9966620 in the *TTC39C* gene, reached genome-wide significance (Table 2) with a *P* value of 4.13×10^{-8} and odds ratio of 1.95 (Confidence interval: 1.53-2.47, Figure 1). This finding was supported by the nearby SNPs (rs7243626 and rs7240470) which also showed encouraging *P* values ($P=5.64 \times 10^{-8}$ and 8.05×10^{-7} , respectively). We calculated the LD among these SNPs using our dataset and these SNPs are highly correlated ($R^2 > 0.8$) (Supplementary Table S3). The regional plot around the top SNPs is shown in the Figure 2. The Q-Q plot of the association

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3 180 results is shown in the Supplementary Figure S2.
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6 181 We have moderate power for this GWAS study. Calculated by CaTS, we have 80% power
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8 182 based on 469 cases and 1,374 controls, assuming a minor disease allele frequency of 0.25, a
9
10 183 genotypic relative risk of 1.49, a prevalence of diabetic maculopathy with decreased visual
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12 184 acuity in the diabetic population of 25%, and significance level of 5×10^{-8} (26).
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15 185 We also ran a GWAS on the phenotype of DM (regardless of visual acuity status, 1,240 cases)
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17 186 against the controls used in this study. The result did not show GWAS significance
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19 187 (Supplementary Table S4).
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24 189 **DISCUSSION**

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26 190 In Scotland, patients with diabetes are invited to attend a free annual retinal screening service.
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29 191 This screening service aims to identify diabetic eye complications at an early stage to prevent or
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31 192 delay subsequent visual loss. During the screening, if an eye falls within the diagnosis criteria of
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33 193 diabetic retinopathy or DM (Supplementary Table S2), then the relevant information will be
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35 194 recorded in the e-health linked records. The status of macula is then categorised and recorded
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38 195 as either: no maculopathy, observable maculopathy, or referable maculopathy. Diabetic
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40 196 retinopathy is further classified and recorded based on the Scottish diabetic retinopathy grading
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42 197 scheme (27). Because of the lack of more detailed classification info, it was not possible to
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45 198 phenotype the subtypes of DM (as outlined by The Royal College of Ophthalmologists
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47 199 guidelines) when using population e-health linked records such as GoDARTS (6), Decreased
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49 200 visual acuity is a well-recognised symptom associated with maculopathy. 22% of diabetic
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51 201 patients with decreased visual acuity have maculopathy, compared to 1% in diabetic patients
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54 202 with normal visual acuity (28). It is also estimated that 6 months after laser treatment, the visual
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3 203 acuity of 34% of DM patients became worse, 22% became better, and 44% remained the same
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5 204 (29). These statistics were similar for DM patients who did not undergo laser treatment. In this
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8 205 study, 37.8% of patients (469/1240) who had DM also had a decreased visual acuity, which is
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10 206 only slightly higher than the above post-laser treatment statistics, however this could be
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12 207 attributed by the fact that we had a longer follow-up period. A national UK audit also found that
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15 208 a poor baseline visual acuity was associated with a poorer visual prognosis in DM patients (30).
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17 209 In this study, we defined the cases as DM with decreased visual acuity based on the hypothesis
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19 210 that these patients might share common genetic components. We tried to identify the genetic
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21 211 factors which contribute to decreased visual acuity among DM patients. Though this narrow
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24 212 definition will reduce case numbers and the study power, it allows us to generate a more
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26 213 homogeneous case population. A similar approach has been applied when we were defining
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28 214 diabetic neuropathic pain (the cases should not only have evidence of pain provided by
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31 215 prescription records, but also positive evidence of neuropathy provided by a medical test) (31).
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33 216 We also acknowledge that 'decreased visual acuity' in the cases was defined as any visual loss,
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35 217 including both major and minor visual loss, which may be an additional confounding factor.
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38 218 Because there is no literature which indicates if diabetic retinopathy and DM share the same
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40 219 genetic components, we also excluded individuals with diabetic retinopathy with and without
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42 220 DM from the controls to maintain a homogeneous population as we wanted to exclude genetic
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44 221 influence from diabetic retinopathy. The duration of diabetes is an important risk factor for many
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47 222 diabetic complications. However, the GoDARTS dataset does not include a high-quality record
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49 223 of this information. Hence, duration of diabetes therefore could not be adjusted for in our
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51 224 analysis. We also noticed that there were statistical differences of some covariates between
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54 225 cases and controls. For example, the mean HbA1c value was higher in cases than controls,
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3 226 which suggest that the controls may have had better controlled diabetes and/or a longer
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5 227 duration of diabetes (32).
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8 228 We have identified the SNP rs9966620, which has achieved GWAS significance ($P=4.13 \times 10^{-8}$,
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10 229 Odds ratio=1.95). This was supported by 2 nearby SNPs which showed encouraging P values.
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12 230 The 3 SNPs are located in the intronic areas with no clear functional roles. The SNP cluster was
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15 231 in the tetratricopeptide repeat domain 39C gene (*TTC39C*), which is a protein-coding gene
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17 232 located on the long arm of chromosome 18. The *TTC39C* gene is expressed in the eyes, and
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19 233 encodes a protein named as TTC39C (33). However, the physiological functions of both the
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21 234 gene and the protein are not known. The protein contains a relatively well-characterized
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24 235 structural motif called the tetratricopeptide repeat (TPR). The TPR is known to be involved in
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26 236 cell cycle regulation, mediating protein-protein interactions, assisting in protein folding and
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28 237 translocation, assembly of multi-protein complexes. The TPR structure also shows flexibility in
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31 238 the mediation of biological activities (34,35). In particular, it has been proposed that this
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33 239 structure plays a role in anaphase: the stage of mitosis when replicated chromosomes are split
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35 240 and the daughter chromatids are moved to opposite poles of the cell (36). Anaphase has been
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38 241 suggested to play an essential role in regulating cell fate of the vertebrate retina (37). Some
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40 242 studies have suggested that central macular thickness, a highly heritable trait with no specific
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42 243 gene identified, is associated with visual acuity (38,39). However, this parameter was not
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45 244 recorded our e-health linked records and so it is still unknown if the *TTC39C* gene is linked with
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47 245 central macular thickness. It has been suggested that the *TTC39C* protein interacts with the
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49 246 protein from the heat shock protein family B (small) member 1 (*HSPB1*) gene (40). *HSPB1* is
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51 247 reported to be upregulated in the rat retina upon optic nerve injury (41). The *TTC39C* protein
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54 248 has 2 paralogs: *TTC39A* and *TTC39B*, although their functions are not clear. Next to *TTC39C*
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are the laminin subunit alpha 3 gene (*LAMA3*) and the calcium binding tyrosine phosphorylation regulated gene (*CABYR*). Of note, the protein of *LAMA3* is laminin subunit alpha-3 and laminins are essential for formation and function of the basement membrane and have additional functions in regulating cell migration and mechanical signal transduction, which suggests it is a possible candidate gene for diabetic maculopathy (42).

We also investigated the top SNPs suggested by other candidate gene studies for diabetic macular oedema. However, none reached statistically significant *P*-values: rs2070744 (*P*=0.286, *NOS3*), rs429358 (*P*=0.59, *APOE*), rs7412 (*P*= 0.35, *APOE*), rs2010963 (*P*=0.30, *VEGFA*), rs17697515 (*P*=0.47, *VEGFC*). Rs2910164 (MicroRNA-146a) was not present in our datasets.

There are limitations in using population level e-health records, especially in differentiating age-related maculopathy from diabetic maculopathy as this information (as well as lens opacity) is not always included. However, the prevalence of age-related maculopathy in a diabetic population is typically low, indicating that this limitation may have had a lessened effect on the study (43). As visual deterioration is associated with foveal oedema, further investigation in a dataset with Optical Coherence Tomography (OCT) data characterising foveal oedema would be valuable to further investigate the findings. A better phenotyping approach of DM for the purposes of research is also required.

In conclusion, we propose that the *TTC39C* gene is associated with DM with decreased visual acuity in a Scottish diabetic cohort using a GWAS approach. Replication studies and functional studies will help to confirm its role in DM.

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12 276 **DISCLOSURE**
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15 277 All authors declare no financial interests or benefit.
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17 278
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38 287 **REFERENCES**
39

- 40 288 1. Cummings TJ. Ophthalmic Pathology. Ophthalmic Pathology. New York, NY: Springer
41
42 289 New York; 2013. 113–132 p.
43
44
45 290 2. Bourne RRA, Stevens GA, White RA, Smith JL, Flaxman SR, Price H, Jonas JB, Keeffe
46
47 291 J, Leasher J, Naidoo K, et al. Causes of vision loss worldwide, 1990-2010: a systematic
48
49 292 analysis. Lancet Glob Heal. 2013 Dec;1(6):e339-49.
50
51 293 doi:10.1016/S2214-109X(13)70113-X. Cited in Pub Med; PMID:25104599.
52
53
54 294 3. Jonas JB, Bourne RRA, White RA, Flaxman SR, Keeffe J, Leasher J, Naidoo K,
55
56
57
58
59

- Pesudovs K, Price H, Wong TY, et al. Visual impairment and blindness due to macular diseases globally: a systematic review and meta-analysis. *Am J Ophthalmol*. 2014 Oct;158(4):808–15. doi:10.1016/j.ajo.2014.06.012. Cited in Pub Med; PMID:24973605.
4. Köberlein J, Beifus K, Schaffert C, Finger RP. The economic burden of visual impairment and blindness: a systematic review. *BMJ Open*. 2013 Nov 7;3(11):e003471. doi:10.1136/bmjopen-2013-003471. Cited in Pub Med; PMID:24202057.
5. Minassian DC, Owens DR, Reidy A. Prevalence of diabetic macular oedema and related health and social care resource use in England. *Br J Ophthalmol*. 2012 Mar;96(3):345–9. doi:10.1136/bjo.2011.204040. Cited in Pub Med; PMID:21602478.
6. Ghanchi F, Diabetic Retinopathy Guidelines Working Group. The Royal College of Ophthalmologists' clinical guidelines for diabetic retinopathy: a summary. *Eye (Lond)*. 2013 Feb 11;27(2):285–7. doi:10.1038/eye.2012.287. Cited in Pub Med; PMID:23306724.
7. Zander E. Maculopathy in patients with diabetes mellitus type 1 and type 2: associations with risk factors. *Br J Ophthalmol*. 2000 Aug 1;84(8):871–6. doi:10.1136/bjo.84.8.871. Cited in Pub Med; PMID:10906094.
8. Leese G. Longitudinal study examining the risk factors for proliferative retinopathy and maculopathy in type-I diabetes: The Royal College of Physicians of Edinburgh Diabetes Register Group. *Eye (Lond)*. 2004 Aug 30;18(8):814–20. doi:10.1038/sj.eye.6701337. Cited in Pub Med; PMID:14752505.
9. Davis MD, Fisher MR, Gangnon RE, Barton F, Aiello LM, Chew EY, Ferris FL, Knatterud GL. Risk factors for high-risk proliferative diabetic retinopathy and severe visual loss: Early Treatment Diabetic Retinopathy Study Report #18. *Invest Ophthalmol Vis Sci*. 1998

1
2
3 318 Feb;39(2):233–52. Cited in Pub Med; PMID:9477980.
4
5 319 10. Awata T, Neda T, Iizuka H, Kurihara S, Ohkubo T, Takata N, Osaki M, Watanabe M,
6
7 Nakashima Y, Sawa T, et al. Endothelial nitric oxide synthase gene is associated with
8 320 diabetic macular edema in type 2 diabetes. Diabetes Care. 2004 Sep;27(9):2184–90.
9
10 321 Cited in Pub Med; PMID:15333482.
11
12 322
13
14 323 11. Awata T, Kurihara S, Takata N, Neda T, Iizuka H, Ohkubo T, Osaki M, Watanabe M,
15
16 Nakashima Y, Inukai K, et al. Functional VEGF C-634G polymorphism is associated with
17 324 development of diabetic macular edema and correlated with macular retinal thickness in
18
19 325 type 2 diabetes. Biochem Biophys Res Commun. 2005 Aug 5;333(3):679–85.
20
21 326 doi:10.1016/j.bbrc.2005.05.167. Cited in Pub Med; PMID:15963467.
22
23 327
24
25 328 12. Kaidonis G, Burdon KP, Gillies MC, Abhary S, Essex RW, Chang JH, Pal B, Pefkianaki
26
27 M, Daniell M, Lake S, et al. Common Sequence Variation in the VEGFC Gene Is
28
29 329 Associated with Diabetic Retinopathy and Diabetic Macular Edema. Ophthalmology.
30
31 330 2015 Sep;122(9):1828–36. doi:10.1016/j.ophtha.2015.05.004. Cited in Pub Med;
32
33 331 PMID:26072347.
34
35 332
36
37 333 13. Kaidonis G, Gillies MC, Abhary S, Liu E, Essex RW, Chang JH, Pal B, Sivaprasad S,
38
39 Pefkianaki M, Daniell M, et al. A single-nucleotide polymorphism in the MicroRNA-146a
40 334 gene is associated with diabetic nephropathy and sight-threatening diabetic retinopathy in
41
42 335 Caucasian patients. Acta Diabetol. 2016 Aug 21;53(4):643–50.
43
44 336 doi:10.1007/s00592-016-0850-4. Cited in Pub Med; PMID:26997512.
45
46 337
47
48 338 14. Meng W, Hebert H, Palmer C. A genome-wide association study suggests that the
49
50 NADPH Oxidase 4 (NOX4) gene is associated with severe diabetic retinopathy in a
51
52 339 Scottish diabetic population. Acta Ophthalmol. 2017 Sep;95.
53
54 340
55
56
57
58
59
60

doi:10.1111/j.1755-3768.2017.02182.

15. Meng W, Shah KP, Pollack S, Toppila I, Hebert HL, McCarthy MI, Groop L, Ahlqvist E, Lyssenko V, Agardh E, et al. A genome-wide association study suggests new evidence for an association of the NADPH Oxidase 4 (NOX4) gene with severe diabetic retinopathy in type 2 diabetes. *Acta Ophthalmol.* 2018 Nov;96(7):e811–9. doi:10.1111/aos.13769. Cited in Pub Med; PMID:30178632.
16. Pollack S, Igo RP, Jensen RA, Christiansen M, Li X, Cheng C-Y, Ng MCY, Smith A V., Rossin EJ, Segrè A V., et al. Multiethnic Genome-Wide Association Study of Diabetic Retinopathy Using Liability Threshold Modeling of Duration of Diabetes and Glycemic Control. *Diabetes.* 2019 Feb;68(2):441–56. doi:10.2337/db18-0567. Cited in Pub Med; PMID:30487263.
17. Awata T, Yamashita H, Kurihara S, Morita-Ohkubo T, Miyashita Y, Katayama S, Mori K, Yoneya S, Kohda M, Okazaki Y, et al. A genome-wide association study for diabetic retinopathy in a Japanese population: potential association with a long intergenic non-coding RNA. Maeda S, editor. *PLoS One.* 2014 Nov 3;9(11):e111715. doi:10.1371/journal.pone.0111715. Cited in Pub Med; PMID:25364816.
18. Graham PS, Kaidonis G, Abhary S, Gillies MC, Daniell M, Essex RW, Chang JH, Lake SR, Pal B, Jenkins AJ, et al. Genome-wide association studies for diabetic macular edema and proliferative diabetic retinopathy. *BMC Med Genet.* 2018 Dec 8;19(1):71. doi:10.1186/s12881-018-0587-8. Cited in Pub Med; PMID:29739359.
19. GoDARTS and UKPDS Diabetes Pharmacogenetics Study Group, Wellcome Trust Case Control Consortium 2, Zhou K, Bellenguez C, Spencer CCA, Bennett AJ, Coleman RL, Tavendale R, Hawley SA, Donnelly LA, et al. Common variants near ATM are associated

1
2
3 364 with glycemic response to metformin in type 2 diabetes. *Nat Genet.* 2011 Feb
4
5 365 26;43(2):117–20. doi:10.1038/ng.735. Cited in Pub Med; PMID:21186350.
6
7
8 366 20. Fagerholm E, Ahlqvist E, Forsblom C, Sandholm N, Syreeni A, Parkkonen M, McKnight
9
10 367 AJ, Tarnow L, Maxwell AP, Parving H-H, et al. SNP in the genome-wide association study
11
12 368 hotspot on chromosome 9p21 confers susceptibility to diabetic nephropathy in type 1
13
14
15 369 diabetes. *Diabetologia.* 2012 Sep;55(9):2386–93. doi:10.1007/s00125-012-2587-0. Cited
16
17 370 in Pub Med; PMID:22643932.
18
19 371 21. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method
20
21 372 for the next generation of genome-wide association studies. Schork NJ, editor. *PLoS*
22
23
24 373 *Genet.* 2009 Jun 19;5(6):e1000529. doi:10.1371/journal.pgen.1000529. Cited in Pub
25
26 374 Med; PMID:19543373.
27
28 375 22. Delaneau O, Marchini J, Zagury J-F. A linear complexity phasing method for thousands of
29
30
31 376 genomes. *Nat Methods.* 2011 Dec 4;9(2):179–81. doi:10.1038/nmeth.1785. Cited in Pub
32
33 377 Med; PMID:22138821.
34
35 378 23. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, Maller J, Sklar P,
36
37
38 379 de Bakker PIW, Daly MJ, et al. PLINK: a tool set for whole-genome association and
39
40 380 population-based linkage analyses. *Am J Hum Genet.* 2007 Sep;81(3):559–75.
41
42 381 doi:10.1086/519795. Cited in Pub Med; PMID:17701901.
43
44
45 382 24. Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and
46
47 383 annotation of genetic associations with FUMA. *Nat Commun.* 2017 Dec;8(1):1826.
48
49 384 doi:10.1038/s41467-017-01261-5. Cited in Pub Med; PMID:29184056.
50
51
52 385 25. Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, Boehnke M,
53
54 386 Abecasis GR, Willer CJ. LocusZoom: regional visualization of genome-wide association
55
56
57
58
59
60

- scan results. *Bioinformatics*. 2010 Sep 15;26(18):2336–7. doi:10.1093/bioinformatics/btq419. Cited in Pub Med; PMID:20634204.
26. Skol AD, Scott LJ, Abecasis GR, Boehnke M. Joint analysis is more efficient than replication-based analysis for two-stage genome-wide association studies. *Nat Genet*. 2006 Feb 15;38(2):209–13. doi:10.1038/ng1706. Cited in Pub Med; PMID:16415888.
27. Liu Y, Wang M, Morris AD, Doney ASF, Leese GP, Pearson ER, Palmer CNA. Glycemic exposure and blood pressure influencing progression and remission of diabetic retinopathy: a longitudinal cohort study in GoDARTS. *Diabetes Care*. 2013 Dec 1;36(12):3979–84. doi:10.2337/dc12-2392. Cited in Pub Med; PMID:24170761.
28. Corcoran JS, Moore K, Agarawal OP, Edgar DF, Yudkin JS. Visual acuity screening for diabetic maculopathy. *Pract Diabetes Int*. 1985 Nov;2(6):30–2. doi:10.1002/pdi.1960020610.
29. Ang GS, Vusirikala B, Mukherji S, Ram ARR. Visual acuity and diabetic maculopathy. *Ann Ophthalmol (Skokie)*. 2006;38(4):305–10. Cited in Pub Med; PMID:17726217.
30. Lee CM, Olk RJ. Modified grid laser photocoagulation for diffuse diabetic macular edema. Long-term visual results. *Ophthalmology*. 1991 Oct;98(10):1594–602. Cited in Pub Med; PMID:1961650.
31. Meng W, Deshmukh HA, van Zuydam NR, Liu Y, Donnelly LA, Zhou K, Wellcome Trust Case Control Consortium 2 (WTCCC2), Surrogate Markers for Micro- and Macro-Vascular Hard Endpoints for Innovative Diabetes Tools (SUMMIT) Study Group, Morris AD, Colhoun HM, et al. A genome-wide association study suggests an association of Chr8p21.3 (GFRA2) with diabetic neuropathic pain. *Eur J Pain*. 2015 Mar;19(3):392–9. doi:10.1002/ejp.560. Cited in Pub Med; PMID:24974787.

1
2
3 410 32. Verma M, Paneri S, Badi P, Raman PG. Effect of increasing duration of diabetes mellitus
4
5 411 type 2 on glycated hemoglobin and insulin sensitivity. Indian J Clin Biochem. 2006
6
7
8 412 Mar;21(1):142–6. doi:10.1007/BF02913083. Cited in Pub Med; PMID:23105586.
9
10 413 33. National Center for Biotechnology Information. EST Profile - Hs.128576 [Internet]. 2019
11
12 414 [cited 2019 Apr 8]. Available from:
13
14
15 415 <https://www.ncbi.nlm.nih.gov/UniGene/ESTProfileViewer.cgi?uglist=Hs.128576>.
16
17 416 34. Allan RK, Ratajczak T. Versatile TPR domains accommodate different modes of target
18
19 417 protein recognition and function. Cell Stress Chaperones. 2011 Jul 9;16(4):353–67.
20
21 418 doi:10.1007/s12192-010-0248-0. Cited in Pub Med; PMID:21153002.
22
23
24 419 35. D'Andrea LD, Regan L. TPR proteins: the versatile helix. Trends Biochem Sci. 2003
25
26 420 Dec;28(12):655–62. doi:10.1016/j.tibs.2003.10.007. Cited in Pub Med; PMID:14659697.
27
28 421 36. Blatch GL, Lässle M. The tetratricopeptide repeat: a structural motif mediating
29
30
31 422 protein-protein interactions. Bioessays. 1999 Nov 11;21(11):932–9.
32
33 423 doi:10.1002/(SICI)1521-1878(199911)21:11<932::AID-BIES5>3.0.CO;2-N. Cited in Pub
34
35 424 Med; PMID:10517866.
36
37
38 425 37. Tibber MS, Kralj-Hans I, Savage J, Mobbs PG, Jeffery G. The orientation and dynamics
39
40 426 of cell division within the plane of the developing vertebrate retina. Eur J Neurosci. 2004
41
42 427 Feb;19(3):497–504. doi:10.1111/j.1460-9568.2004.03172.x. Cited in Pub Med;
43
44 428 PMID:14984400.
45
46
47 429 38. Liew SHM, Gilbert CE, Spector TD, Marshall J, Hammond CJ. The role of heredity in
48
49 430 determining central retinal thickness. Br J Ophthalmol. 2007 Sep 1;91(9):1143–7.
50
51 431 doi:10.1136/bjo.2007.114215. Cited in Pub Med; PMID:17360735.
52
53
54 432 39. Diabetic Retinopathy Clinical Research Network, Browning DJ, Glassman AR, Aiello LP,
55
56
57
58
59
60

- Beck RW, Brown DM, Fong DS, Bressler NM, Danis RP, Kinyoun JL, et al. Relationship between optical coherence tomography-measured central retinal thickness and visual acuity in diabetic macular edema. *Ophthalmology*. 2007 Mar;114(3):525–36. doi:10.1016/j.ophtha.2006.06.052. Cited in Pub Med; PMID:17123615.
40. Orchard S, Ammari M, Aranda B, Breuza L, Briganti L, Broackes-Carter F, Campbell NH, Chavali G, Chen C, Del-Toro N, et al. The MIntAct project--IntAct as a common curation platform for 11 molecular interaction databases. *Nucleic Acids Res*. 2014 Jan;42(Database issue):D358-63. doi:10.1093/nar/gkt1115. Cited in Pub Med; PMID:24234451.
41. Schmidt T, Fischer D, Andreadaki A, Bartelt-Kirbach B, Golenhofen N. Induction and phosphorylation of the small heat shock proteins HspB1/Hsp25 and HspB5/ α B-crystallin in the rat retina upon optic nerve injury. *Cell Stress Chaperones*. 2016 Jan 16;21(1):167–78. doi:10.1007/s12192-015-0650-8. Cited in Pub Med; PMID:26475352.
42. National Center for Biotechnology Information. LAMA3 laminin subunit alpha 3 [Homo sapiens (human)] - Gene - NCBI [Internet]. 2019 [cited 2019 Apr 8]. Available from: <https://www.ncbi.nlm.nih.gov/gene/3909>.
43. Borrone R, Saravia M, Bar D. Age related maculopathy and diabetes. *Eur J Ophthalmol*. 18(6):949–54. Cited in Pub Med; PMID:18988167.

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TABLES

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FIGURE CAPTIONS

- Figure 1. Manhattan plot of the GWAS on diabetic maculopathy with decreased visual acuity (469 cases and 1,374 controls).
- Figure 2. Regional plot around top SNPs of the TTC39C gene area.
- Supplementary Figure S1. Workflow of the GWAS on diabetic maculopathy with decreased visual acuity in GoDARTS.
- Supplementary Figure S2. Q-Q plot compared expected and observed $-\text{Log}_{10}(\text{P})$ values.

Table 1. Covariates information between cases and controls.

	Cases (mean+SD)	Controls (mean+SD)	<i>P</i>
Age , y	67.20±9.50	65.27±10.39	0.0003
Sex , n	192/277 (m/f)	630/744 (m/f)	0.068
BMI , kg/m ²	31.25±4.99	31.77±5.84	0.082
Cholesterol, mmol/L	4.36±0.85	4.40±0.88	0.372
Triglycerides, mmol/L	2.07±1.10	2.29±1.30	0.002
HDL, mmol/L	1.37±0.36	1.33±0.34	0.018
LDL, mmol/L	2.09±0.67	2.10±0.72	0.870
HbA1c, %	8.00±1.46	7.27±1.20	<0.0001

Cases= 469

Controls= 1,374

SD: standard deviation

BMI: body mass index

HDL: high-density lipoprotein

LDL: low-density lipoprotein

Table 2. SNPs of the GWAS on diabetic maculopathy with decreased visual acuity ($p<1.0\times10^{-6}$).

SNPID	Chromosome position (GRCh37)	Gene	Minor Allele	MAF in cases: controls	<i>P</i> value (no adjustment)	<i>P</i> value	OR \pm SE
rs9966620	18:21680735	<i>TTC39C</i>	A	16.33%:9.64%	7.23×10^{-8}	4.13×10^{-8}	1.95 \pm 0.12
rs7243626	18:21679950	<i>TTC39C</i>	T	16.17%:9.51%	8.84×10^{-8}	5.64×10^{-8}	1.95 \pm 0.12
rs7240470	18:21681516	<i>TTC39C</i>	T	16.86%:10.33%	3.20×10^{-7}	8.05×10^{-7}	1.80 \pm 0.12
rs11706588	3:126448513	<i>CHCHD6</i>	C	20.98%:13.05%	6.99×10^{-7}	4.43×10^{-7}	1.89 \pm 0.13
rs11718070	3:126448560	<i>CHCHD6</i>	T	20.98%:13.09%	8.01×10^{-7}	5.13×10^{-7}	1.88 \pm 0.13

SNP: single nucleotide polymorphism

MAF: minor allele frequency

OR: odds ratio

SE: standard error

TTC39C: tetratricopeptide repeat domain 39C

CHCHD6: coiled-coil-helix-coiled-coil-helix domain containing 6

rs11706588 and rs11718080 are included in the table for readers' interest although the *P* values of these 2 SNPs did not reach GWAS significance.

Listed covariates in the Table 1 were used for adjustments.

Table S1. The definition of visual acuity in GoDARTS.

Visual acuity coding	Description
1	6/4
2	6/5
3	6/6
4	6/9
5	6/12
6	6/18
7	6/24
8	6/36
9	6/60
10	3/60
11	Counting fingers
12	Hand movements
13	Perception of Light
14	No perception of Light

Visual acuity of 6/12 means a testing eye can see at six metres while an eye with normal vision can see at 12 metres away.

Individuals with coding 11 to 14 were removed from study.

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Table S2. Maculopathy and retinopathy grades for the diabetic retinal screen.

MACULOPATHY GRADING		
Maculopathy	Description	Outcome
M0 (No maculopathy)	No features ≤ 2 disc diameters from the centre of the fovea, enough to classify it as M1 or M2 from definitions below	Rescreen in 12 months
M1 (Observable)	Any hard exudates within a radius of > 1 but ≤ 2 disc diameters of the fovea centre	Rescreen in 6 months or refer to ophthalmology if not feasible
M2 (Referable)	Any hard exudates or blot haemorrhages within a distance of ≤ 1 disc diameter of the fovea centre.	Refer to ophthalmology (these patients will not definitely receive immediate laser treatment and may be kept under surveillance
RETINOPATHY GRADING		
Retinopathy	Description	Outcome
R0 (No visible retinopathy)	No diabetic retinopathy anywhere	Rescreen in 12 month
R1 (Mild)	Background Diabetic Retinopathy (BDR) – Mild Presence of any one of these: Dot haemorrhages, micro-aneurysms, hard exudates, cotton-wool spots, blot haemorrhages, flames shaped haemorrhages	Rescreen in 12 month
R2 (Observable Background)	Background Diabetic Retinopathy (BDR) – Observable Four(4) or more blot haemorrhages in one hemi-field*	Rescreen in 6 month
R3 (Referable Background)	Background Diabetic Retinopathy (BDR) - Referable Presence of any of the following: <ul style="list-style-type: none">• Four(4) or more blot haemorrhages in both superior and inferior hemifields• Venous beading• IrMA	Refer to ophthalmology;
R4 (Proliferative)	Proliferative Diabetic Retinopathy (PDR) Presence of active new vessels or Vitreous haemorrhage	Refer to ophthalmology; Patients are likely to receive laser treatment or other intervention
R6 (Inadequate)	Not adequately visualized; Retina not sufficiently visible for assessment	Technical failure; Patients for alternative screening examination.
IrMA, Intraretinal Microvascular Abnormality. * The hemifields, Superior and Inferior, are demarcated by a line through the fovea centre and optic disc.		

Source: The Scottish Diabetic Retinopathy Grading Scheme 2007 v1.0.¹⁶

Table S3. Linkage disequilibrium among 3 SNPs.

SNPID	rs7243626	rs9966620	rs7240470
rs7243626	1	1	0.84
rs9966620	1	1	0.85
rs7240470	0.84	0.85	1

SNP: single nucleotide polymorphism

The values in the table indicate R-square values.

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Table S4. The top SNPs for the GWAS on the phenotypes of DM (regardless of visual acuity, 1,240 cases) against the controls (1,374 samples).

Chromosome	SNP	Positions	Test	NMISS	OR	SE	L95	U95	<i>P</i>
1	rs3818329	192627760	ADD	2324	1.865	0.1202	1.474	2.361	2.12x10 ⁻⁷
3	rs12629668	147209179	ADD	2061	1.397	0.06611	1.228	1.591	4.15x10 ⁻⁷
6	rs117482282	165511471	ADD	2361	3.079	0.2224	1.991	4.762	4.27x10 ⁻⁷
13	rs1149833	50750876	ADD	2557	0.747	0.05813	0.6666	0.8372	5.23x10 ⁻⁷
2	rs1406230	29583321	ADD	1943	0.6923	0.07334	0.5996	0.7994	5.35x10 ⁻⁷
1	rs2296022	192627124	ADD	2324	1.799	0.1183	1.427	2.269	6.92x10 ⁻⁷
16	rs35498131	9120809	ADD	1493	0.5596	0.117	0.445	0.7038	6.94x10 ⁻⁷
7	rs140306040	62321151	ADD	2171	0.509	0.1365	0.3895	0.6651	7.47x10 ⁻⁷
7	rs73121760	62313911	ADD	2155	0.5131	0.135	0.3938	0.6685	7.71x10 ⁻⁷
16	rs34300094	9120719	ADD	1487	0.5589	0.1184	0.4431	0.7049	8.98x10 ⁻⁷
2	rs34954281	152225877	ADD	2021	0.6739	0.08045	0.5756	0.789	9.34x10 ⁻⁷

SNP: single nucleotide polymorphism

NMISS: number of individuals for the logistic regression analysis of a specific SNP

OR: odds ratio

SE: standard error

ADD: additive model

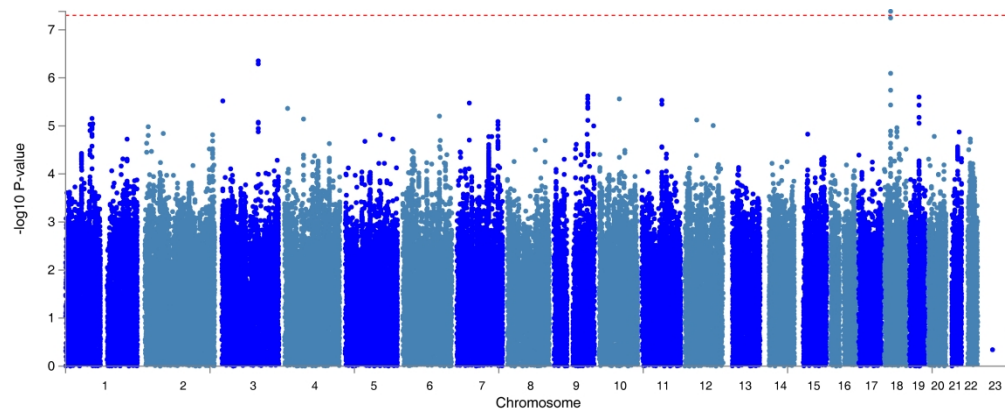


Figure 1. Manhattan plot of the GWAS on diabetic maculopathy with decreased visual acuity (469 cases and 1,374 controls).

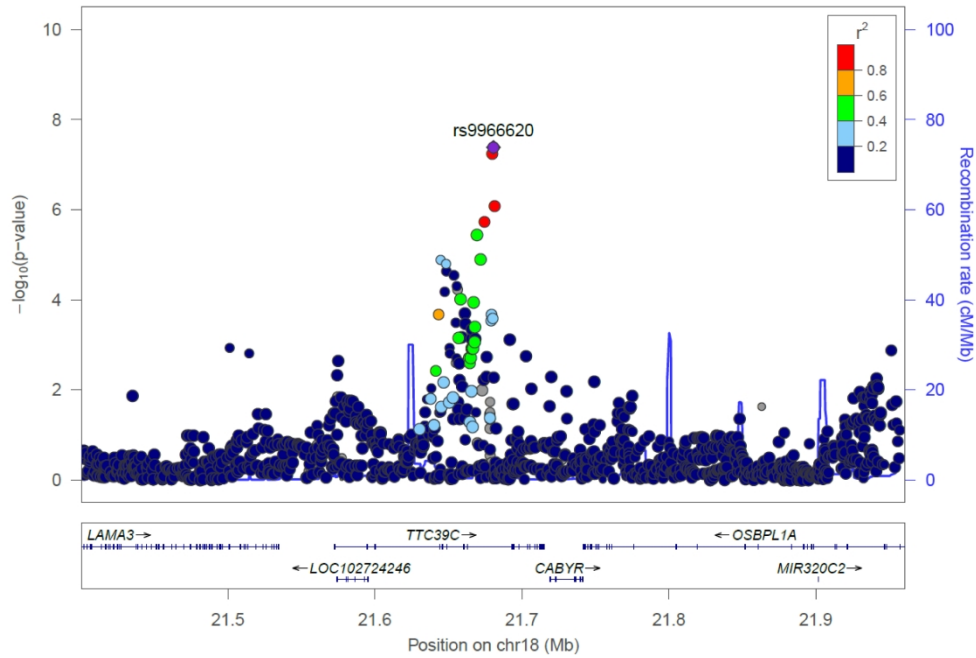


Figure 2. Regional plot around top SNPs of the TTC39C gene area.


1. 3673 individuals were genotyped by WTCCC2 using Affymetrix SNP6.0
3254 individuals were genotyped by SUMMIT using Illumina OmniExpress
 2. Perform imputation using SHAPEIT and IMPUTE2, adapt imputation quality control ($r^2 > 0.3$)
 3. Extract imputed genotypes of cases and controls according to their definitions
 4. Merge, detect population stratification, remove relatives and perform routine quality control
 5. Obtain cleaned datasets including 469 cases and 1,374 controls in PLINK format ($\lambda = 1.00$)
 6. Logistic regression analyses with covariates in PLINK
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Figure S1. Workflow of the GWAS on diabetic maculopathy with decreased visual acuity in GoDARTS.

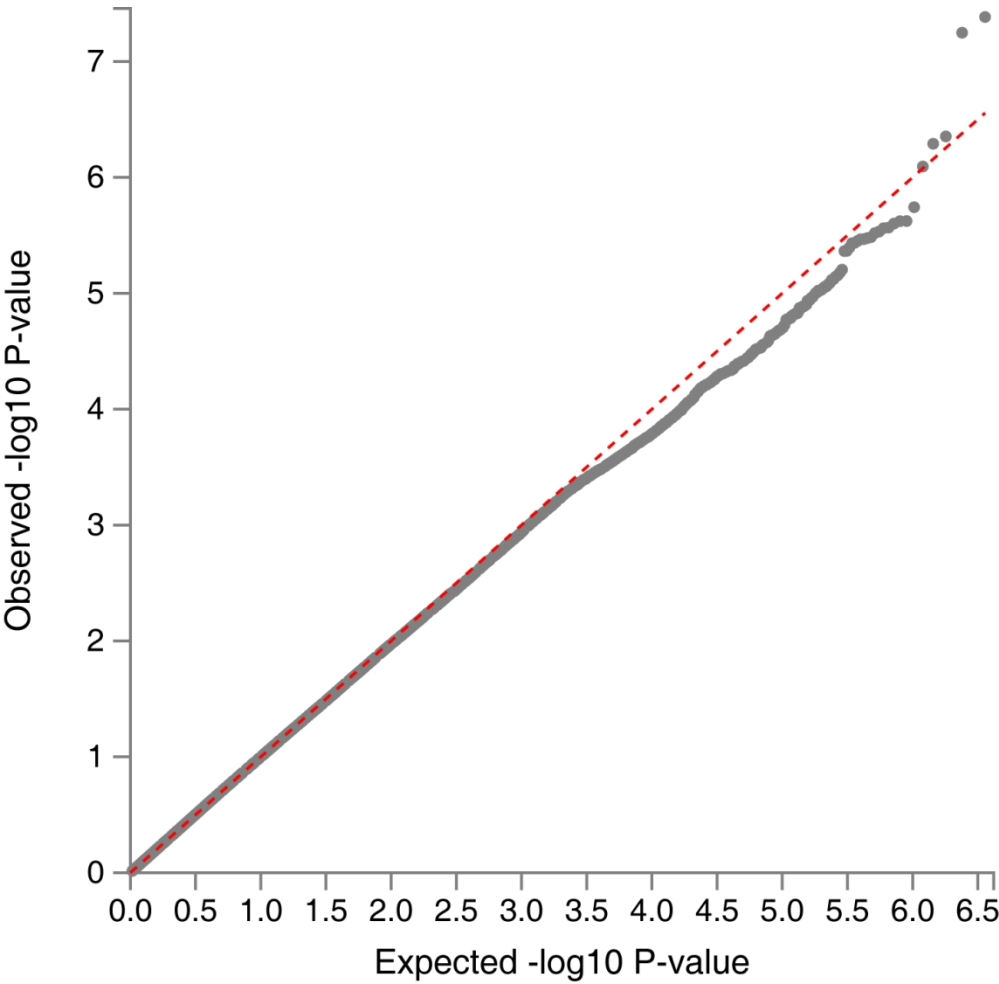


Figure S2. Q-Q plot compared expected and observed $-\log_{10}(P)$ values.